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## Output O.T3.1

### *Ex situ* gene stocks of Danube sturgeons

Project co-funded by European Union funds (ERDF, IPA)





# Output O.T3.1 – *Ex situ* gene stocks of Danube sturgeons

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## 1. Introduction

Sturgeon populations represent an important historic, economic and natural heritage of the Danube River Basin and are prone to serve as indicators of good ecological status. Their products such as meat and caviar are well known rendering these species part of tradition and culture of the Danube Region. Sturgeons are also sensitive animals, which indicate environmental pressures. As flagship-species, they are valuable indicators for the health of river ecosystems. However, their populations suffered substantial losses from decades of overfishing, habitat fragmentation and pollution (Guti 2006) mainly in the late 20<sup>th</sup> and early 21<sup>st</sup> centuries. Fragmentation of the migratory route creates an ecological barrier restricting the habitat to be used and isolating population segments while permitting only one-way gene flow. The strongest effects upon longitudinal connectivity are resulting from the Iron Gate Dams and the Gabčíkovo Dam but multiple smaller dams fragment the Upper Danube and the tributaries throughout the catchment (Friedrich et al. 2019). Furthermore, habitat degradation is a potent threat, as side arms and sections with slower current velocities historically provided feeding and nursing grounds for migratory species, but drainage and river control measures largely eliminated these sections. Furthermore, new threats of invasive species and global climate change are likely to exert even more pressures on these species and their protection is more urgently required than ever before.

The goals of habitat protection, restoration of migratory routes, restocking events and different educational and strategic approaches have been outlined by the ICPDR and its Sturgeon 2020 program. This program was followed by the network of national and international stakeholders in the “Danube Sturgeon Task Force”. The DSTF is linked to the ICPDR and the Priority Area 6 of the EU Strategy for the Danube Region (Biodiversity and Landscape Diversity), which harmonizes sectoral policies between environmental protection and economic requirements.

The DTP2-038-2.3 – MEASURES project is part of the Interreg Danube Transnational Programme and was designed to promote the functionality of ecological corridors in the Danube River Basin. The stability of migratory fish populations requires long-term restoration of river connectivity however the populations decrease at such speed, that immediate short-term protection is necessary to ensure the survival of the species in question. The MEASURES project targets both time-scales and reviews the knowledge and practical solutions that serve these issues. Although multiple species of migratory fish are

accounted for in the MEASURES project, sturgeons are especially targeted by additional measures.

One of the most straightforward actions for protecting endangered species, is to create *ex situ* stocks of representative entities of populations in question in a safe and controlled environment to maintain ecologically viable population sizes in such facilities. Thus, providing a backup for the dwindling natural populations under controlled conditions. These serve as a basis for their reintroduction once the conditions of the natural habitats are sufficiently suitable. This method has significant value for protecting individuals, but also provides the basis for controlled propagation and restocking. During these occasions large quantities of genetically controlled, viable individuals are released into the natural habitats to support or establish wild populations. These actions not only strengthen natural populations, but provide a visible platform to engage with stakeholders and public audience about the issues of migratory fish species and eco-corridors.

Of the six sturgeon species, native to the Danube River Basin (*Huso huso*, *Acipenser gueldenstaedtii*, *A. nudiiventris*, *A. ruthenus*, *A. stellatus*, *A. sturio*), the sterlet (*A. ruthenus*) and the Russian sturgeon (*Acipenser gueldenstaedti*) were selected in the frame of the MEASURES project to strengthen their natural populations through pilot restocking and genetic analysis of wild specimens and captive broodstock.

## **2. Establishing *ex situ* stocks of Danube sturgeons**

Due to the endangered nature of the Danube sturgeon species, sufficiently sized and diverse *ex situ* stocks, representing the full width of plasticity of the wild populations, maintained under controlled conditions are key for their survival. These *ex situ* stocks provide the basis for annual artificial reproductions, comparison of stocks by genetic profiling and potential for releasing vulnerable fish species in their natural environment. For the maintenance of *ex situ* stocks, it is vital to collect wild specimens. These individuals can contribute genetically to the diversity of the already existing *ex situ* populations when they recruit into the broodstock. It is also important that for restocking purposes, wild fish in the *ex situ* stock must originate from the same region where the planned release is to take place (genetic integrity). Collecting wild specimens is followed by biometric and genetic analysis as well. These actions help to ensure a proper phenotypic and genetic background for the selected breeders in the gene stock, and consequently, long-term genetic stability of both *ex situ* population as well as the released individuals.

## **3. Sterlet *ex situ* stock**

NAIK HAKI has a long history in genetic *ex situ* conservation management. An adequate population size of the sterlet is available for reproduction and releases or for supplementary expansion of the *ex situ* stock.

The *ex situ* sturgeon broodstock represents great value due to the long-term efforts that have been required to establish it. As such its maintenance requires above average safety (provision more careful rearing conditions than in aquaculture production; low stocking,

proper feeding, optimal water quality variables, etc.) and tracking measures. For tracking purposes, each individual has its own PIT tag to identify and trace the *ex situ* stock at the individual level. The genetically analyzed sterlet broodstock (not the whole *ex situ* stock) in this study originates from the municipal district of Leányfalu, Ercsi, Baja, Mindszent and Kisköre. The first three of these locations can be found at the Hungarian sections of the Middle-Danube, the last two are from the Tisza River, which is the largest tributary of Danube in Hungary.

### **3.1 Broodstock reinforcement and gene bank maintenance**

In project MEASURES, NAIK-HAKI attempted to capture sterlet broodstock from the Middle-Danube section in 2019 and 2020, but only when additional help of external fishermen was utilized in 2020, the attempt was successful. The institute managed to capture additional 10 sterlets (originate from Leányfalu, Danube) prior the spawning season and transfer them to NAIK-HAKI into controlled conditions, where they successfully survived and adapted to the different environment in the upcoming critical months. The sterlets had 1.73 kg average weight per individual, which is a size generally smaller than the generic broodstock population, and only 2 of them were sufficiently mature to be used in the propagation procedure in 2020. This catch was used to expand the existing sterlet *ex situ* stock at NAIK-HAKI. Including the new 10 Danubian individuals, NAIK-HAKI genotyped 144 sterlet brooders (3,03 kg average weight per individual) in 2020.

#### Reproduction planning and theoretical considerations

In the MEASURES project, there were two artificial propagation events at NAIK-HAKI which were planned to increase the sterlet stocks available. For reproduction purposes, in an ideal case we choose 15 “ready to spawn” (which were not reproduced in the previous year) specimens from each sex to ensure the appropriate genetic diversity in the next generation.

#### Reproduction preparations

From those 30 selected individuals at least 20 (10 females and 10 males) are designated for propagation to contribute to the next generation. The selected broodstock are transferred from their maintenance pond to hatchery tanks 4-5 m<sup>3</sup> each with the same temperature, 6-8 mg/L dissolved oxygen concentration and pH 7.5-8. If the external water temperature is lower than 14°C then slow raising in water temperature (max 0.5°C/day) has to be applied.

#### Timing and induction of reproduction

For induced reproduction we use analogues of mammalian luteinizing hormone-releasing hormone, which is the des-Gly10(D-Ala6)-LH-RH Ethylamid. This hormone has been successfully used in several sturgeon species and the dose at 40 µg/kg for females can be used to induce final oocyte maturation in sterlet.

On 14°C 34-46 hours after a successful induction eggs are collected by an incision of the caudal section of the oviducts. Milt is obtained from the males by pressing the abdominal wall. The stickiness of the fertilized eggs is eliminated by a talc-water solution (500g talc/10L water). Eggs are incubated in Zug jars, 300-400 g fertilized, treated (not sticky) eggs can be loaded in a Zug jar with the volume of 7L. The water flow rate during incubation is around 1.5-2.5 L/min. Polyvinylpyrrolidone (PVP) Iodine treatment was used to control fungus in

every 12 hours during incubation with 10 mL in each Zug jar for 10 minutes without water flow. The hatching starts about five days after fertilization at 16-16.5°C.

During hatching larvae are transferred to larger tanks (preferably circle tank) with shallow water depth (30-50 cm) and lower water flow (5L/min). The onset of exogenous feeding takes place around 9 dph (day of post hatch) when the melanin plug being evacuated from the gastrointestinal tract.

In the first days of active feeding larvae are fed with *Artemia nauplii* or fine-chopped Chironomid worms.

The nursing largely depends on the method and efficiency of weaning to artificial diets. The weaning of sterlet batch for release starts around 25 dph. The duration of weaning is usually 3-4 days, when the quantity of natural food is gradually decreased and that of artificial feed is increased. Some of these feeds have been adapted from salmonid culture however, special starter feeds for sturgeons are also manufactured. The protein content of the applied feeds is between 45-50% with particle size of 0.2-0.8 mm. For releasing purposes the adequate portion of natural food is further provided at least once a day, parallel to the artificial diet.

### 3.2. Sterlet reproduction in 2019

In total 27 sexually matured individuals were selected and transported to the hatchery. From the selected individuals 16 were females and 9 were males. After the hormone induction, eggs from 9 ovulated females were successfully stripped and milt from 5 males were collected (**Table 1**). The motility of the milt was checked and only the motile sperm was mixed prior to fertilization. In total 7,790 g of dry eggs were obtained. After successful fertilization the average fertilization rate was 49.2%.

**Table 1.** General description and notes for sterlet reproduction in 2019

Sex	Species	PIT tag ID	01/04/2019	09/04/2019	Notes
			Weight (kg)	Reproduction	
♀	Sterlet broodstock	0415D5026D	5,10	✓	1060 g eggs
	Sterlet broodstock	0415D527CA	3,40	×	no eggs
	Sterlet broodstock	452F5B5148	2,90	×	no eggs
	Sterlet broodstock	0415D4C256	2,95	✓	529 g eggs
	Sterlet broodstock	0415D4F4F1/4AA64	4,75	×	no eggs
	Sterlet broodstock	0415D51BD4	4,60	×	no eggs
	Sterlet broodstock	0415D521F8	4,70	×	no eggs
	Sterlet broodstock	0415D50028	2,70	×	No eggs
	Sterlet broodstock	4349763139	5,00	✓	810 g eggs
	Sterlet broodstock	0415D52286	2,10	✓	460 g eggs
	Sterlet broodstock	0415D5270D	4,00	✓	948 g eggs
	Sterlet broodstock	0415D4FF49	3,60	✓	990 g eggs/died
	Sterlet broodstock	0415D51ABF	4,10	✓	1400 g eggs
	Sterlet broodstock	0415D51143	2,40	✓	663 g eggs
	Sterlet broodstock	0415D52C4C	3,80	✓	930 g eggs
♀♂	Sterlet broodstock	4461467344	2,50	×	hermaphrodite
♂	Sterlet broodstock	0415D5074D	1,80	✓	
	Sterlet broodstock	0415D508F7	4,50	×	no sperm
	Sterlet broodstock	0415D50826	4,40	✓	
	Sterlet broodstock	0415D4A984	2,40	×	no sperm
	Sterlet broodstock	434857336A	3,50	×	no sperm
	Sterlet broodstock	0415D529B6	4,00	✓	
	Sterlet broodstock	0415D52CC5	3,25	×	no sperm
	Sterlet broodstock	0415D4A0CB	3,50	×	no sperm
	Sterlet broodstock	0415D5208F/525EF	1,90	✓	
	Sterlet broodstock	4534647709	1,80	×	no sperm
	Sterlet broodstock	0415D500ED	1,30	✓	

### 3.3. Sterlet reproduction in 2020

In 2020, only 2 of the captured 10 sterlets from Leányfalu proved to be matured enough for reproduction. For reproduction we selected 15 “ready to spawn” females from the gene bank and 2 Danubian wild females, in case of males we selected 12 from the gene bank and got 8 from the Danube. We successfully stripped eggs from 12 females and one of them was

Danubian wild. In the case of males we got sperm from 12 individuals and one of them was Danubian wild (**Table 2**). The motility of the sperm was checked and only the motile sperm was mixed prior to fertilization. In total 8,004 g of dry eggs were obtained. After fertilization the average fertilization rate was 48.4%.

**Table 2.** General description and notes for sterlet reproduction in 2020

Sex	Species	PIT tag ID	02/04/2020	06/04/2020	Notes
			Weight (kg)	Reproduction	
♀	<b>Danube wild sterlet</b>	0415D4C642	3,65	×	no eggs
	<b>Danube wild sterlet</b>	0415D4C9EB	3,60	✓	569 g eggs
	Sterlet broodstock	0415D4FFB0	5,00	✓	1034 g eggs
	Sterlet broodstock	0415D528D2	4,20	✓	784 g eggs
	Sterlet broodstock	0415D52753	3,95	✓	970 g eggs
	Sterlet broodstock	0415D516FB	3,75	✓	755 g eggs
	Sterlet broodstock	0415D51646	3,75	✓	1060 g eggs
	Sterlet broodstock	0415D52362	3,35	✓	370 g eggs
	Sterlet broodstock	0415D51ADD	3,15	×	no eggs
	Sterlet broodstock	0415D50190	3,15	✓	736 g eggs
	Sterlet broodstock	0415D5237C	3,10	✓	486 g eggs
	Sterlet broodstock	0415D51143	3,10	✓	614 g eggs
	Sterlet broodstock	0415D4AD5C	3,10	✓	264 g eggs
	Sterlet broodstock	0415D5128E	2,75	×	no eggs
	Sterlet broodstock	0415D5208B	2,55	×	no eggs
	Sterlet broodstock	0415D4B2A1	2,50	×	no eggs
	♂	<b>Danube wild sterlet</b>	0415D4BC62	1,90	×
<b>Danube wild sterlet</b>		0415D4B72E	1,30	×	no sperm
<b>Danube wild sterlet</b>		0415D4BB98	1,25	✓	
<b>Danube wild sterlet</b>		0415D4C4AA	1,20	×	no sperm
<b>Danube wild sterlet</b>		0415D4B615	1,15	×	no sperm
<b>Danube wild sterlet</b>		0415D4B0F4	1,15	×	no sperm
<b>Danube wild sterlet</b>		0415D50FB1	1,15	×	no sperm
<b>Danube wild sterlet</b>		0415D51F79	0,90	×	no sperm
Sterlet broodstock		0415D4F7E1	5,60	✓	
Sterlet broodstock		0415D52778	4,00	✓	milted two times
Sterlet broodstock		0415D5119C	3,35	×	no sperm
Sterlet broodstock		0415D502CF	2,95	✓	
Sterlet broodstock		0415D4FE29	2,90	✓	
Sterlet broodstock		4534647709	2,70	✓	
Sterlet broodstock		0415D515A8	2,70	✓	
Sterlet broodstock		0415D49E39	2,70	✓	
Sterlet broodstock		0415D4DF51	2,65	✓	
Sterlet broodstock	0415D500ED	2,40	✓		
Sterlet broodstock	0415D4B063	2,10	✓		
Sterlet broodstock	0415D4B118	1,80	✓		



### 3.4. Juvenile reinforcements

Resulting from the two sterlet reproductions conducted in MEASURES at NAIK-HAKI, the broodstock of total 146 individuals was supplemented with juveniles. From 2019, 1,700 juveniles were reared for more than a year, of which 1,500 were used for the sterlet restocking event in 2020 (Baja, Hungary). The remaining juveniles (192 individuals, considering survival) achieved 0.92 kg average body weight until 2020. The reproduction event from 2020 provided an additional 700 juveniles, which have reached a 0.15 kg average body weight until 2020.

The cooperation of the MEASURES project and the LIFE-Sterlet project provided the background for an additional transfer of Danubian sterlet fry and juveniles stemming from wild parents in Austria from BOKU to NAIK-HAKI in 2020. Juveniles hatched in 2018 (2 individuals, 0.4 kg average weight), in 2019 (25 individuals, 0.16 kg average weight) and fry from 2020 (100 pcs, 0.05 kg average weight) were delivered in the summer 2020. Since then, survival was 75%, with 2 individuals from the 2018 batch, 24 individuals from the 2019 batch and 77 individuals of the 2020 batch having survived.

### 3.5. Genetic profiling of sterlet broodstock

For strengthening gene conservation and restocking events, it is important to evaluate the genetic background of the caught wild broodstock. For the analysis to detect variability and genotype microsatellite markers are used. This procedure helps to determine allele-richness and the genetic similarity of individuals from the Danube and its most important tributary, river Tisza.

For this purpose, we assessed the genotype of 144 individuals with 12 microsatellite (DNA) markers from a wild population. The collected fish originated from river Tisza, Kisköre and Mindszent (22 pcs together) and from three sections of river Danube (Leányfalu (D1) – 47 pcs, Ercsi (D2) – 30 pcs, Baja (D3) – 30 pcs and 15 fish originated from the existing *ex situ* stock, this fish was propagated in 2019 and 2020.). The fish were marked with PIT tags, to collect genetic data on the individual level. For the analysis we took samples from the tissue on the caudal fin. DNA isolation was conducted using the E.N.Z.A. kit tissue DNA (Omega Bio-tek Inc.) method (following the instructions set by the producer) and 20-30 mg tissue samples. The quantity and quality of the extracted DNA were assessed using NanoDrop 2000c spectrophotometer (Thermo Scientific, USA). We used 100 ng DNA per sample for the PCR reaction and 12 microsatellite markers for the genetic analysis. During the setting of PCR reaction, we relied on the instructions set by the producer (Multiplex PCR Plus Kit, Qiagen) and optimized the specific settings. The size and quality of the amplified products were assessed on 1.5% agarose and detected the fragments using ABI 3130 sequencing (Applied biosystems, USA). The allele data produced from the electropherograms were identified by the Gene Mapper (Applied Biosystem) software. The specific allele data from the 144 analyzed individuals are presented in **Annex/Table A1**. The specific markers and allele numbers in the table represent the results of the genotyping.

The standard population genetic calculations were delivered by the GenALEx 6.5 (Peakall, Smouse 2012) statistical software package. This was followed by the identification of the genetic distance between populations. Additionally, the assessment of heterozygosity-ratio allowed us to make conclusions regarding the actual genetic structure of the populations. As a result of the genotyping, we detected 118 alleles on the 12 loci. The number of alleles per loci are presented in **Table 3**.

**Table 3** The number of alleles for each locus

LOCUS	ALLELE NUMBER
AN_20	15
AFUG_41	24
ARU_26	8
SPL_163	16
AFUG_51	6
ARU_12	3
ARU_13	21
LS_68	17
ARU_19	2
AOXD_161	12
ARU-50	7
ARU_18	4
TOTAL	135

We analyzed the genetic variability and allelic pattern of populations by loci (**Table 4.**) and calculated averages for populations (**Table 5.**). Different parameters were included in the analysis: number of differentiable genotypes, the relationship between expected and experienced heterozygosity, fixation index. The allele numbers for loci varied between 2 and 24 (11.25 in average).

**Table 4** Characteristics of the genetic variability and allelic pattern: number of differentiable genotypes (N), number of alleles (Na), experienced (Ho) and expected (He) heterozygosity, fixation index (F).

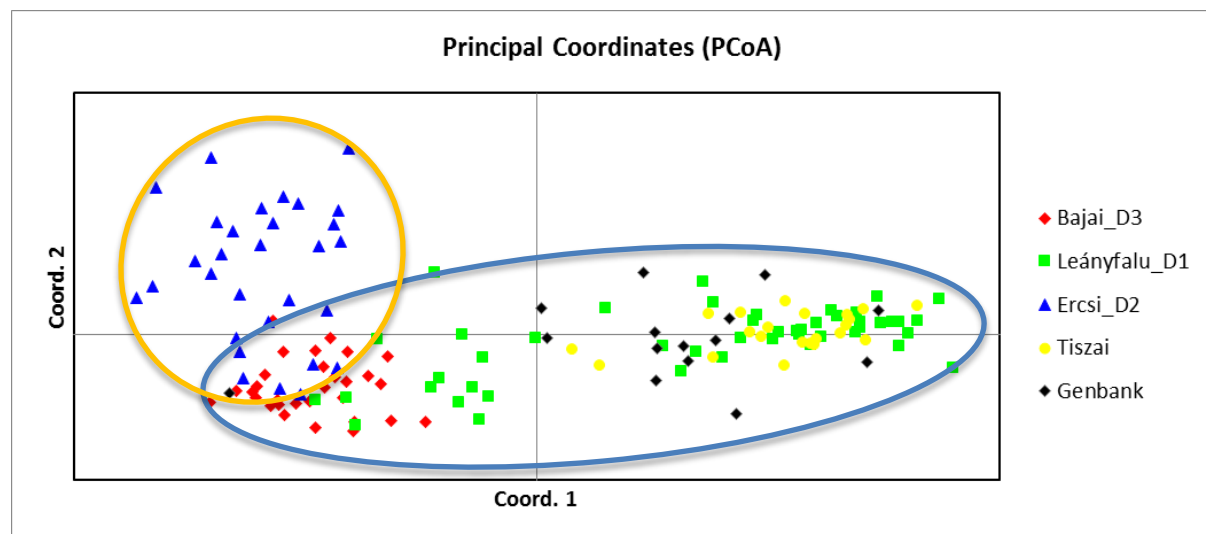
Population		AN_20	AfuG_41	ARU_26	Spl_163	AFUG_51	ARU_12	Aru_13	LS_68	Aru_19	AOXD_161	Aru-50	Aru_18
Bajai_D3	N	30	30	30	22	30	30	30	26	30	30	30	30
	Na	9	12	6	9	1	2	7	7	2	5	3	2
	Ho	0,73	0,57	0,47	0,64	0,00	0,97	0,63	0,35	0,37	0,60	0,03	0,23
	He	0,81	0,89	0,66	0,87	0,00	0,50	0,67	0,82	0,34	0,58	0,49	0,21
	F	0,10	0,36	0,29	0,27	0,00	-0,94	0,06	0,58	-0,08	-0,04	0,93	-0,13
Leányfalu_D1	N	43	44	47	45	46	45	46	40	47	47	47	46
	Na	10	17	3	14	5	2	15	16	2	6	6	4
	Ho	0,84	0,82	0,15	0,76	0,17	0,76	0,76	0,43	0,06	0,64	0,89	0,24
	He	0,83	0,91	0,46	0,82	0,27	0,48	0,88	0,92	0,10	0,57	0,63	0,22
	F	-0,01	0,10	0,68	0,08	0,36	-0,57	0,13	0,54	0,37	-0,12	-0,41	-0,11
Ercsi_D2	N	28	28	30	27	30	30	29	29	30	30	30	29
	Na	9	15	3	11	3	2	7	8	2	7	3	2
	Ho	0,64	0,96	0,57	0,22	0,37	0,90	0,55	0,38	0,37	0,80	0,07	0,10
	He	0,79	0,85	0,42	0,82	0,31	0,50	0,65	0,63	0,47	0,78	0,13	0,10
	F	0,19	-0,14	-0,37	0,73	-0,20	-0,80	0,15	0,40	0,22	-0,03	0,47	-0,05
Tiszai	N	20	20	21	21	21	21	21	21	22	22	22	22
	Na	7	9	2	9	4	3	15	12	2	7	3	2
	Ho	1,00	0,80	0,10	0,71	0,24	0,90	0,95	0,62	0,09	0,82	0,86	0,09
	He	0,79	0,84	0,09	0,68	0,22	0,52	0,87	0,89	0,09	0,63	0,54	0,09
	F	-0,27	0,04	-0,05	-0,05	-0,09	-0,75	-0,10	0,30	-0,05	-0,30	-0,60	-0,05
Genbank	N	13	13	14	14	15	14	15	15	15	15	14	15
	Na	10	11	2	6	2	3	8	12	2	7	3	3
	Ho	0,92	0,92	0,36	0,64	0,00	0,86	0,67	0,80	0,27	0,73	0,71	0,27
	He	0,85	0,86	0,38	0,77	0,12	0,56	0,75	0,88	0,23	0,67	0,54	0,24
	F	-0,09	-0,07	0,05	0,16	1,00	-0,53	0,11	0,09	-0,15	-0,10	-0,32	-0,12

Most alleles were detected in the group from Leányfalu, which was followed by populations from Tisza, Ercsi, Genbank and Baja.

**Table 5** Genetic variability and averages of the allelic pattern

Population	Bajai_D3	Leányfalu_	Ercsi_D2	Tiszai	Genbank
Na	5,42	<b>8,33</b>	6	6,25	5,75
Na Freq. >= 5%	4,08	5	3,42	3,25	4
I	1,19	1,39	1,12	1,15	1,22
No. Private Alleles	0,5	<b>0,92</b>	0,67	0,42	<b>0,33</b>
No. LComm Alleles (<=50%)	0,42	1,17	0,5	0,75	<b>0,58</b>
He	0,57	0,59	0,54	0,52	0,57
Ho	0,47	0,54	0,49	0,6	0,6
F	0,13	0,09	0,05	-0,16	0

Values of heterozygosity were used to calculate the F ratio (fixation index), which provides information about the differentiation process of populations. Assuming absolute fixation, that is, the presence of only homozygotes, this value is 1. The average values of fixation index per populations were always close to zero and in the population from Tisza this value was even negative. These results indicate the dominance of heterozygotes in the populations. Among the populations analyzed, the genetic variability between individuals is relatively high and there are no signs for inbreeding. The values from the expected and experienced heterozygosity and their connection also provide important insight regarding the populations. The average expected heterozygosity (He) for the four populations were between 0.52 and 0.57, while the experienced (Ho) values were between 0.47 and 0.61. Comparing population-wise, these values stand close to each other. In the population from Tisza, the averaged experienced heterozygosity exceeded the expected value, which also supports an excess of heterozygotes.

**Figure 1.** Results of the PCoA test of GenALEx

The genetic differences were calculated using the GENALEX software and we graphically compared the genetic linkages and differences between populations. The results of the PCoA (Principle Coordinate Analysis) analysis are presented in **Figure 1**. The applied procedure assigns characteristic genetic profiles for the groups and then enlists the specific individuals into the most likely groups, based on their calculated genetic value. This can be seen in the figure, where the presentation of individual values helps to explore the genetic relations between the specific groups. The genetic background of the analyzed individuals from the five populations with the applied markers separates two main groups (circled in the PCoA

graph). The Leányfalu, Baja, Tisza and Genebank stocks belong to one of them (blue circle) and the other group contains the Ercsi group (orange circle). The Leányfalu, Baja, Tisza and Genebank populations overlap in population genetic point of view. The genetic pattern of Ercsi stock might refer to the result of previous restocking activities from captured stocks.

#### **4. Russian sturgeon gene stocks**

Russian sturgeons are virtually extinct in the Middle-Danube region and NAIK-HAKI has only two Danubian (17.0 kg average weight) and 14 Caspian Russian sturgeons (19.2 kg average weight) as broodstock. The NAIK HAKI additionally purchased 100 pcs Russian sturgeon juveniles with Danube origin from Romania (from 2014 reproduction), since then 95 juveniles survived (1.96 kg average weight) as a potential new generation for broodstock enhancement.

Unfortunately, Russian sturgeon broodstock was not available for the MEASURES project to purchase or obtain from the natural environment. However, the project partners managed to purchase 3,000 fertilized Russian sturgeon eggs with the contribution of five brooders of Danube origin in Romania. The purchase was conducted by the Danube Delta National Institute for Research and Development (DDNI) in early 2020, and the eggs were transported by the Institute of Biodiversity and Ecosystem Research - Bulgarian Academy of Sciences (IBER-BAS) to the University of Natural Resources and Life Sciences, Vienna (BOKU) for hatching. The eggs were incubated and the juveniles reared in Danube water in close cooperation with the LIFE-Sterlet project (LIFE14/NAT/AT/000057). Roughly 1,800 larvae started to feed after pre-larval development. On agreement between project partners, 500 of them were transported to NAIK-HAKI in July for further gene bank enhancement and rearing. Of these 471 survived until September, 2020 and have 0.11 kg average weight. More than 500 stayed in Austria to split the risk and as a basis for further conservation projects. They have an average weight of 0.23 kg as of the end of September. Additionally a little over 300 specimens with around 15 cm TL (total length) have been stocked in the Nationalpark Donauauen in July.

#### **5. Conclusions**

Fishing of sterlet from natural waters is experienced more difficult than before and the acquisition of Russian sturgeon individuals is even more troublesome, the MEASURES project successfully increased *ex situ* stocks of both of these species. The new sterlet brooders partially could support the reproduction event in NAIK-HAKI, 2020. Although Russian sturgeon broodstock was not available for purchase, 3,000 fertilized eggs were used by the project partners for gene bank enhancement at BOKU and at NAIK-HAKI as well. The fail-safe recirculation systems provide high protection for fry and juveniles, and the PIT tagging helps to track older individuals to monitor their development closely. The transportation of endangered fish species in MEASURES helped to share the needed infrastructural capacity

for mass rearing and also supported the transnational network of multiple, *ex situ* gene banks.

## 6. References

Friedrich, T., Reinartz, R., & Gessner, J. (2019). Sturgeon re-introduction in the Upper and Middle Danube River Basin. *Journal of Applied Ichthyology*, 35(5), 1059-1068.

Guti, G. (2006). Past and present status of sturgeons in Hungary. In *Proceedings of the 36th International Conference of IAD. Austrian Committee Danube Research/IAD, Vienna* (pp. 143-147).

Peakall, R., & Smouse, P. E. (2012). GenAEx 6.5: genetic analysis in Excel. Population genetic software for teaching and researchdan update. *Bioinformatics* 28, 2537e2539.



